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Chapter I

Introduction to prokaryotic metabolism and physiology

The biosphere has been shaped both by physical events and by interactions with the organisms that occupy it. Among living organisms, prokaryotes are much more metabolically diverse than eukaryotes and can also thrive under a variety of extreme conditions where eukaryotes cannot. This is possible because of the wealth of genes, metabolic pathways and molecular processes that are unique to prokaryotic cells. For this reason, prokaryotes are very important in the cycling of elements, including carbon, nitrogen, sulfur and phosphorus, as well as metals and metalloids such as copper, mercury, selenium, arsenic and chromium. Prokaryotes are important not only for shaping the biosphere, but are also involved in the health of plants and animals including humans. Disease-causing bacteria have been a major concern in microbiology from the dawn of the science, while recent developments in 'microbiome' research reveal paramount roles in the well-being of higher plants and animals. A full understanding of the complex biological phenomena that occur in the biosphere therefore requires a deep knowledge of the unique biological processes that occur in this vast prokaryotic world.

After publication in 1995 of the first full DNA sequence of a free-living bacterium, *Haemophilus influenzae*, whole genome sequences of thousands of prokaryotes have now been determined and many others are currently being sequenced (see https://gold.jgi.doe.gov/ and https://www.ncbi.nlm.nih.gov/guide/gen omes-maps/). Our knowledge of the whole genome profoundly influences all aspects of microbiology. Determination of entire genome sequences, however, is only a first step in fully understanding the properties of an organism and its interactions with the environment in which it lives. The functions encoded by these sequences need to be elucidated to give biochemical, physiological and ecological meaning to the information. Furthermore, sequence analysis indicates that the biological functions of substantial portions of complete genomes are so far unknown. Defining the role of each gene in the complex cellular metabolic network is a formidable task. In addition, genomes contain hundreds to thousands of genes, many of which encode multiple proteins that interact and function together as multicomponent systems for accomplishing specific cellular processes. The products of many genes are often co-regulated in complex signal transduction networks, and understanding how the genome functions as a whole presents an even greater challenge. It is also known that for a significant proportion of metabolic activities, no representative genes have been identified across all organisms, such activities being termed as 'orphan' to indicate they are not currently assigned to any gene. This also represents a major future challenge and will require both computational and experimental approaches.

It is widely accepted that less than 1 per cent of prokaryotes have been cultivated in pure culture under laboratory conditions. This is also

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2 INTRODUCTION TO PROKARYOTIC METABOLISM AND PHYSIOLOGY

true of the majority of species associated with higher organisms, including humans, which have not been isolated in pure culture but play important roles in the well-being of the host. Development of new sequencing techniques has allowed us to obtain genomic information from the multitudes of unculturable prokaryotic species and complex microbial populations that exist in nature. Such information might provide a basis for the development of new cultivation techniques. Elucidation of the function of unknown genes through a better understanding of biochemistry and physiology could ultimately result in a fuller understanding of the complex biological phenomena occurring in the biosphere.

Unlike multicellular eukaryotes, individual cells of unicellular prokaryotes are more exposed to the continuously changing environment, and have evolved unique structures and metabolic processes to survive under such conditions. Chapter 2 describes the main aspects of the composition and structure of prokaryotic cells.

Life can be defined as a reproduction process using materials available from the environment according to the genetic information possessed by the organism. Utilization of the materials available in the environment necessitates transport into cells that are separated from the environment by a membrane. Chapter 3 outlines transport mechanisms, not only for intracellular entry of nutrients, but also for excretion of materials including extracellular enzymes and materials that form cell surface structures.

Many prokaryotes, including *Escherichia coli*, can grow in a simple mineral salts medium containing glucose as the sole organic compound. Glucose is metabolized through glycolytic pathways and the tricarboxylic acid (TCA) cycle, supplying all carbon skeletons, energy in the form of ATP and reducing equivalents in the form of NADPH for growth and reproduction. Glycolysis is described in Chapter 4 with emphasis on the reverse reactions of the EMP pathway and on prokaryote-specific metabolic pathways. When substrates other than glucose are used, parts of the metabolic pathways are employed in either forward or reverse directions. Chapter 5 describes the TCA cycle and related metabolic pathways, and energy transduction mechanisms. Chapter 6 describes the biosynthetic metabolic processes that utilize carbon skeletons, ATP and NADPH, the production of which is discussed in the previous chapters. These chapters summarize the biochemistry of central metabolism that is employed by prokaryotes to enable growth on a glucose-mineral salts medium.

The next five chapters describe metabolism in some of the various trophic variations found in prokaryotes. These are the use of organic compounds other than glucose as carbon and energy sources (Chapter 7), anaerobic fermentation (Chapter 8), anaerobic respiratory processes (Chapter 9), chemolithotrophy (Chapter 10) and photosynthesis (Chapter 11). Some of these metabolic processes are prokaryote specific, while others are found in both prokaryotes and eukaryotes.

Prokaryotes only express a proportion of their genes at any given time, just like eukaryotes. This enables them to grow in the most efficient way under any given conditions. Metabolism is regulated not only through control of gene expression but also by controlling the activity of enzymes. These regulatory mechanisms are discussed in Chapter 12. Finally, the survival of prokaryotic organisms under starvation conditions is discussed in terms of storage materials, resting cell structures and population survival in Chapter 13.

This book has been written as a text for senior students at undergraduate level and postgraduates in microbiology and related subjects. A major proportion of the book has been based on review papers published in various scientific journals including those listed below:

Annual Review of Microbiology Annual Review of Biochemistry Current Opinion in Microbiology FEMS Microbiology Reviews Journal of Bacteriology Microbiology and Molecular Biology Reviews (formerly Microbiology Reviews) Nature Reviews Microbiology Trends in Microbiology

The authors would also like to acknowledge the authors of the books listed below that have been consulted during the preparation of this book. Cambridge University Press & Assessment 978-1-316-62291-9 — Prokaryotic Metabolism and Physiology Byung Hong Kim , Geoffrey Michael Gadd Excerpt More Information

FURTHER READING 3

- Caldwell, D. R. (2000). *Microbial Physiology and Metabolism*, 2nd edn. Belm, CA: Star Publishing Co.
- Dawes, D. A. (1986). *Microbial Energetics*. Glasgow: Blackie.
- Dawes, I. W. & Sutherland, I. W. (1992). Microbial Physiology, 2nd edn. Basic Microbiology Series, 4. Oxford: Blackwell.
- Gottschalk, G. (1986). Bacterial Metabolism, 2nd edn. New York: Springer-Verlag.
- Ingraham, J. L., Maaloe, O. & Neidhardt, F. C. (1983). *Growth of the Bacterial Cell.* Sunderland, MA: Sinauer Associates Inc.
- Mandelstam, J., McQuillin, K. & Dawes, I. (1982). *Biochemistry of Bacterial Growth*, 3rd edn. Oxford: Blackwell.

Further Reading

Note this section contains key references only. Additional recommended references are available at www.cambridge.org/ProkaryoticMetabolism.

General

- Downs, D. M. (2006). Understanding microbial metabolism. *Annual Review of Microbiology* **60**, 533–559.
- Solden, L., Lloyd, K. & Wrighton, K. (2016). The bright side of microbial dark matter: lessons learned from the uncultivated majority. *Current Opinion in Microbiology* **31**, 217–226.

Diversity

- Achtman, M. & Wagner, M. (2008). Microbial diversity and the genetic nature of microbial species. *Nature Reviews Microbiology* **6**, 431–440.
- Bertin, P. N., Medigue, C. & Normand, P. (2008). Advances in environmental genomics: towards an integrated view of micro-organisms and ecosystems. *Microbiology* **154**, 347–359.
- Fernandez, L. A. (2005). Exploring prokaryotic diversity: there are other molecular worlds. *Molecular Microbiology* 55, 5–15.

Ecology and Geomicrobiology

Ehrlich, H. L., Newman, D. K. & Kappler, A. (2015). Ehrlich's Geomicrobiology, 6th edn. Boca Raton, FL, USA: CRC Press.

- Moat, A. G., Foster, J. W. & Spector, M. P. (2002). *Microbial Physiology*, 4th edn. New York: Wiley.
- Neidhardt, F. C. & Curtiss, R. (eds.) (1996). Escherichia coli and Salmonella: Cellular and Molecular Biology, 2nd edn. Washington, DC: ASM Press.
- Neidhardt, F. C., Ingraham, J. L. & Schaechter, M. (1990). *Physiology of the Bacterial Cell: A Molecular Approach*. Sunderland, MA: Sinauer Associates Inc.
- Stanier, R. J., Ingraham, J. L., Wheelis, M. K. & Painter, P. R. (1986). *The Microbial World*, 5th edn. Upper Saddle River, NJ: Prentice-Hall.
- White, D. (2000). *The Physiology and Biochemistry of Prokaryotes*, 2nd edn. Oxford: Oxford University Press.
- Gadd, G. M., Semple, K. T. & Lappin-Scott, H. M. (2005). Micro-organisms and Earth Systems: Advances in Geomicrobiology. Cambridge: Cambridge University Press.
- Konhauser, K. O. (2007). Introduction to Geomicrobiology. Malden, MA, USA: Blackwell Science Ltd.
- Madsen, E. L. (2015). Environmental Microbiology: From Genomes to Biogeochemistry, 2nd edn. New York: Wiley.
- Shively, J. M., English, R. S., Baker, S. H. & Cannon, G. C. (2001). Carbon cycling: the prokaryotic contribution. *Current Opinion in Microbiology* 4, 301–306.
- Vorholt, J. A. (2012). Microbial life in the phyllosphere. Nature Reviews Microbiology **10**, 828–840.

Evolution

- Boucher, Y., Douady, C. J., Papke, R. T., Walsh, D. A., Boudreau, M. E., Nesbo, C. L., Case, R. J. & Doolittle, W. F. (2003). Lateral gene transfer and the origins of prokaryotic groups. *Annual Review of Genetics* 37, 283–328.
- Boyd, E. F., Almagro-Moreno, S. & Parent, M. A. (2009). Genomic islands are dynamic, ancient integrative elements in bacterial evolution. *Trends in Microbiology* **17**, 47–53.
- Koch, A. L. & Silver, S. (2005). The first cell. Advances in Microbial Physiology 50, 227–259.
- van der Meer, J. R. & Sentchilo, V. (2003). Genomic islands and the evolution of catabolic pathways in bacteria. *Current Opinion in Biotechnology* **14**, 248–254.

4 | INTRODUCTION TO PROKARYOTIC METABOLISM AND PHYSIOLOGY

Genomics

- Chun, J. & Rainey, F. A. (2014). Integrating genomics into the taxonomy and systematics of the Bacteria and Archaea. *International Journal of Systematic and Evolutionary Microbiology* **64**, 316–324.
- Francke, C., Siezen, R. J. & Teusink, B. (2005). Reconstructing the metabolic network of a bacterium from its genome. *Trends in Microbiology* **13**, 550–558.
- Loman, N. J. & Pallen, M. J. (2015). Twenty years of bacterial genome sequencing. *Nature Reviews Microbiology* 13, 787–794.
- Medini, D., Serruto, D., Parkhill, J., Relman, D. A., Donati, C., Moxon, R., Falkow, S. & Rappuoli, R. (2008). Microbiology in the post-genomic era. *Nature Reviews Microbiology* 6, 419–430.
- Ward, N. & Fraser, C. M. (2005). How genomics has affected the concept of microbiology. *Current Opinion in Microbiology* **8**, 564–571.

Extreme environments

- Bowers, K. & Wiegel, J. (2011). Temperature and pH optima of extremely halophilic archaea: a mini-review. *Extremophiles* **15**, 119–128.
- Cowan, D. A. (2004). The upper temperature for life where do we draw the line? *Trends in Microbiology* **12**, 58–60.
- Javaux, E. J. (2006). Extreme life on Earth: past, present and possibly beyond. *Research in Microbiology* **157**, 37–48.

Human microbiome

- Bashan, A., Gibson, T. E., Friedman, J., Carey, V. J., Weiss, S. T., Hohmann, E. L. & Liu, Y.-Y. (2016). Universality of human microbial dynamics. *Nature* 534, 259–262.
- Bauer, K. C., Huus, K. E. & Finlay, B. B. (2016). Microbes and the mind: emerging hallmarks of the gut microbiota-brain axis. *Cellular Microbiology* **18**, 632–644.
- Borody, T. J. & Khoruts, A. (2012). Fecal microbiota transplantation and emerging applications. *Nature Reviews Gastroenterology and Hepatology* **9**, 88–96.
- Consortium, T. H. M. P. (2012). Structure, function and diversity of the healthy human microbiome. *Nature* **486**, 207–214.
- Garrett, W. S. (2015). Cancer and the microbiota. *Science* **348**, 80–86.
- Grice, E. A. & Segre, J. A. (2011). The skin microbiome. Nature Reviews Microbiology **9**, 244–253.
- Hooper, L. V., Littman, D. R. & Macpherson, A. J. (2012). Interactions between the microbiota and the immune system. *Science* **336**, 1268–1273.
- O'Toole, P. W. & Jeffery, I. B. (2015). Gut microbiota and aging. *Science* **350**, 1214–1215.
- Sharon, G., Sampson, T. R., Geschwind, D. H. & Mazmanian, S. K. (2016). The central nervous system and the gut microbiome. *Cell* **167**, 915–932.
- Sommer, F., Anderson, J. M., Bharti, R., Raes, J. & Rosenstiel, P. (2017). The resilience of the intestinal microbiota influences health and disease. *Nature Reviews Microbiology* **15**, 630–638.

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Chapter 2

Composition and structure of prokaryotic cells

Like all organisms, microorganisms grow, metabolize and replicate utilizing materials available from the environment. Such materials include those chemical elements required for structural aspects of cellular composition and metabolic activities such as enzyme regulation and redox processes. To understand bacterial metabolism, it is therefore helpful to know the chemical composition of the cell and component structures. This chapter describes the elemental composition and structure of prokaryotic cells, and the kinds of nutrients needed for biosynthesis and energy-yielding metabolism.

2.1 Elemental composition

From over 100 naturally occurring elements, microbial cells generally only contain 12 in significant quantities. These are known as major elements, and are listed in Table 2.1 together with some of their major functions and predominant chemical forms used by microorganisms.

They include elements such as carbon (C), oxygen (O) and hydrogen (H) constituting organic compounds like carbohydrates. Nitrogen (N) is found in microbial cells in proteins, nucleic acids and coenzymes. Sulfur (S) is needed for S-containing amino acids, such as methionine and cysteine, and for various coenzymes. Phosphorus (P) is present in nucleic acids, phospholipids, teichoic acid and nucleotides including NAD(P) and ATP. Potassium is the major inorganic cation (K^+), while chloride (Cl^-) is the major inorganic anion. K⁺ is required as a cofactor for certain enzymes, e.g. pyruvate kinase. Chloride is involved in the energy conservation process utilized by halophilic archaea (Section 11.6). Sodium (Na⁺) participates in several transport and energy transduction processes, and plays a crucial role in microbial growth under alkaline conditions (Section 5.7.4). Magnesium (Mg^{2+}) forms complexes with phosphate groups including those found in nucleic acids, ATP, phospholipids and lipopolysaccharides. Several microbial intracellular enzymes, e.g. monomeric alkaline phosphatase, are calcium dependent. Ferrous and ferric ions play a crucial role in oxidation-reduction reactions as components of electron carriers such as Fe-S proteins and cytochromes.

In addition to these 12 major elements, others are also found in microbial cells as minor elements (Table 2.2). All the metals listed in Table 2.2 are required for specific enzymes. It is interesting to note that the atomic number of tungsten is far higher than that of the other elements, and that this element is only required in rare cases.

In addition to those listed in Table 2.2, some unusual elements are used by microorganisms. Under molybdenum-limited conditions, a vanadium-containing nitrogenase is synthesized in nitrogen-fixing organisms (Section 6.2.1.2). A claim that arsenate can substitute for phosphate in the synthesis of certain macromolecules,

| COMPOSITION AND STRUCTURE OF PROKARYOTIC CELLS

Element	Atomic number	Chemical forms used by microbes	Function
С	6	organic compounds, CO, CO ₂	major constituents of cell material in proteins, nucleic acids, lipids, carbohydrates and others
0	8	organic compounds, CO ₂ , H ₂ O, O ₂	
Н	I	organic compounds, H_2O , H_2	
Ν	7	organic compounds, NH4 ⁺ , NO3 ⁻ , N2	
S	16	organic sulfur compounds, SO_4^{2-} , HS ⁻ , S ⁰ , S ₂ O ₃ ²⁻	proteins, coenzymes
Р	15	HPO ₄ ^{2–} , organophosphate organic phosphonates, phosphite	nucleic acids, phospholipids, teichoic aci coenzymes
К	19	K ⁺	major inorganic cation, compatible solut enzyme cofactor
Mg	12	Mg ²⁺	enzyme cofactor, bound to cell wall, membrane and phosphate esters including nucleic acids and ATP
Ca	20	Ca ²⁺	enzyme cofactor, bound to cell wall
Fe	26	Fe ²⁺ , Fe ³⁺	cytochromes, ferredoxin Fe-S proteins, enzyme cofactor
Na		Na ⁺	involved in transport and energy transduction
Cl	17	CI ⁻	major inorganic anion

including DNA, in a bacterium isolated from an arsenic-rich lake has been extensively debated. A similarly toxic metal, cadmium, is contained in the carbonic anhydrase of a marine diatom. Growth of extremely acidophilic methanotrophic Methylacidiphilum fumariolicum on methanol is strictly dependent on the presence of lanthanides, a group of the rare earth elements (REEs) that includes lanthanum (La), cerium (Ce), praseodymium (Pr) and neodymium (Nd). Lanthanides are an essential cofactor in its methanol dehydrogenase (MDH). This enzyme (XoxF) is different from the MxaF-type MDH, containing calcium as a catalytic cofactor of Methylobacterium extorquens (Section 7.10.2). The XoxF-type MDH is induced by La and Ce in other methylotrophs, Bradyrhizobium sp. and Methylobacterium radiotolerans. Metagenomic

studies have shown that XoxF-type MDH is much more prominent in nature than the MxaF-type enzymes. Silicate can be solubilized by a group of bacteria known as silicate bacteria, including *Bacillus circulans*, but silicon does not appear to have any essential roles in prokaryotic biology.

2.2 Importance of chemical form

2.2.1 Five major elements

The elements listed in Tables 2.1 and 2.2 need to be supplied or be present in the chemical forms that the organisms can use. Carbon is the most abundant element in all living organisms. Prokaryotes are broadly classified according to the carbon sources they use: organotrophs Cambridge University Press & Assessment 978-1-316-62291-9 — Prokaryotic Metabolism and Physiology Byung Hong Kim, Geoffrey Michael Gadd Excerpt More Information

2.2 IMPORTANCE OF CHEMICAL FORM 7

Table 2.2 Minor elements found in microbial cells with their functions and predominant chemical formused by microorganisms.				
Element	Atomic number	Chemical form used by microbes	Function	
Mn	25	Mn ²⁺	superoxide dismutase, photosystem II	
Со	27	Co ²⁺	coenzyme B ₁₂	
Ni	28	Ni ²⁺	hydrogenase, urease	
Cu	29	Cu ²⁺	cytochrome oxidase, oxygenase	
Zn	30	Zn ²⁺	alcohol dehydrogenase, aldolase, alkaline phosphatase, RNA and DNA polymerase, arsenate reductase	
Se	34	SeO ₃ ²⁻	formate dehydrogenase, glycine reductase	
Mo	42	MoO4 ²⁻	nitrogenase, nitrate reductase, formate dehydrogenase, arsenate reductase	
\mathbb{W}	74	WO_4^{2-}	formate dehydrogenase, aldehyde oxidoreductase	

(heterotrophs) use organic compounds as their carbon source while CO_2 is used by lithotrophs (autotrophs). These groups are divided further according to the form of energy they use: chemotrophs (chemoorganotrophs and chemolithotrophs) depend on chemical forms for energy, while phototrophs (photoorganotrophs and photolithotrophs) utilize light energy ('organo' refers to an organic substance while 'litho' refers to an inorganic substance).

Nitrogen sources commonly used by microbes include organic nitrogenous compounds such as amino acids, and inorganic forms such as ammonium and nitrate. Gaseous N_2 can serve as a nitrogen source for nitrogen-fixing prokaryotes. Nitrogen fixation is not known in eukaryotic microorganisms. Some chemolithotrophs can use ammonium as their energy source (electron donor, Section 10.2) while nitrate can be used as an electron acceptor by denitrifiers (Section 9.1).

Sulfate is the most commonly used sulfur source, while other sulfur sources used include organic sulfur compounds, sulfide, elemental sulfur and thiosulfate. Sulfide and sulfur can serve as electron donors in certain chemolithotrophs (Section 10.3), and sulfate and elemental sulfur are used as electron acceptors and are reduced to sulfide by sulfidogens (Section 9.3). Phosphate is the most common phosphorus (P) source used by microorganisms, and many microorganisms use organophosphate. When the phosphate supply is limited, various organophosphonates are used as a P source in microorganisms. Unlike organophosphates that possess C–O–P linkages, phosphonates have direct C–P linkages. These are not only produced in nature as antibiotics (e.g. fosfomycin) and herbicides (e.g. phosphinothricin), but are also chemically synthesized for various commercial applications such as the broad-spectrum herbicide glyphosate. Phosphite is also used as a P source by many marine microorganisms.

2.2.2 Oxygen

Oxygen in cells originates mainly from organic compounds, water or CO_2 . Molecular oxygen (O_2) is seldom used in biosynthetic processes. Some prokaryotes use O_2 as the electron acceptor, but some cannot grow in its presence. Thus, organisms can be grouped according to their reaction with O_2 , into aerobes that require O_2 , facultative anaerobes that use O_2 when it is available but can also grow in its absence, and obligate anaerobes that do not use O_2 . Some obligate anaerobes cannot grow, and die in the presence of O_2 , while others can tolerate it. The former are termed

8 | COMPOSITION AND STRUCTURE OF PROKARYOTIC CELLS

strict anaerobes and the latter aerotolerant anaerobes.

2.2.3 Growth factors

Some organotrophs, such as *Escherichia coli*, can grow in simple media containing glucose and mineral salts, while others, like lactic acid bacteria, require complex media containing various compounds such as vitamins, amino acids and nucleic acid bases. This is because the latter organisms cannot synthesize certain essential cellular materials from only glucose and mineral salts. These required compounds should therefore be supplied in the growth media: such compounds are known as growth factors. Growth factor requirements differ between organisms, with vitamins being the most commonly required growth factors (Table 2.3).

2.3 Structure of microbial cells

Microorganisms are grouped into either prokaryotes or eukaryotes according to their cellular structure. With only a few exceptions, prokaryotic cells do not have subcellular organelles separated from the cytoplasm by phospholipid membranes, such as the eukaryotic nuclear and mitochondrial membranes. Organelles, like the nucleus, mitochondria and endoplasmic reticulum, are only found in eukaryotic cells. The detailed structure of prokaryotic cells is described below.

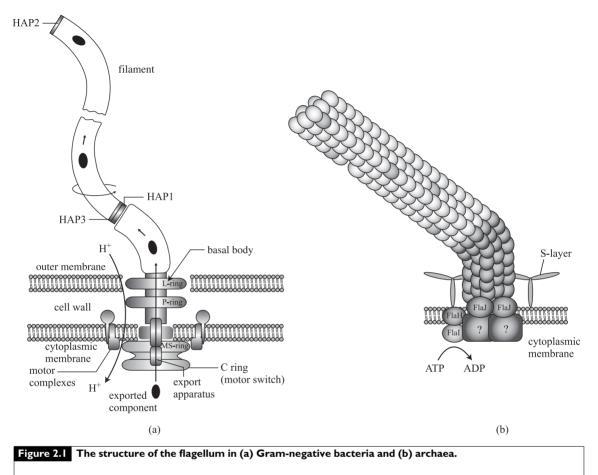
2.3.1 Flagella and pili

Motile prokaryotic cells have an appendage called a flagellum (plural, flagella), involved in motility, and a similar but smaller structure, the fimbria (plural, fimbriae). Fimbriae are not involved in motility and are composed of proteins.

The bacterial flagellum consists of three parts. These are a basal body, a hook and a filament (Figure 2.1). The basal body is embedded in the cytoplasmic membrane and cell surface structure and connected to the filament through the hook. In Gram-negative bacteria the basal body consists of a cytoplasmic membrane ring, a periplasmic ring and an outermembrane ring through which the central rod **Table 2.3**Common growth factors required byprokaryotes and their major functions.

Growth factor	Function
p-aminobenzoate	component of tetrahydrofolate, a one-carbon unit carrier
Biotin	prosthetic group of carboxylase and mutase
Coenzyme M	methyl carrier in methanogenic archaea
Folate	component of tetrahydrofolate
Haemin (Hemin)	precursor of cytochromes and haemoproteins
Lipoate	prosthetic group of 2-keto acid decarboxylase
Nicotinate	precursor of pyridine nucleotides (NAD ⁺ , NADP ⁺)
Pantothenate	precursor of coenzyme A and acyl carrier protein
Pyridoxine	precursor of pyridoxal phosphate
Riboflavin	precursor of flavins (FAD, FMN)
Thiamine	precursor of thiamine pyrophosphate
Vitamin B ₁₂	precursor of coenzyme B_{12}
Vitamin K	precursor of menaquinone

passes. The diameter of the rings can be 20– 50 nm depending on the species. The cytoplasmic ring of the basal body is associated with additional proteins known as the Mot complex. The Mot complex rotates the basal body with the entire flagellum consuming a proton motive force (or sodium motive force). The cytoplasmic membrane ring is therefore believed to function as a motor with the Mot complex. A more detailed description of motility is given in Section 12.2.11. In addition to the Mot complex, the basal body is associated with an export apparatus through which the building blocks of the filament are transported. Cambridge University Press & Assessment 978-1-316-62291-9 — Prokaryotic Metabolism and Physiology Byung Hong Kim, Geoffrey Michael Gadd Excerpt More Information



(Modified from Nature Rev. Microbiol. 6: 466-476, 2008)

(a) Gram-negative bacterial flagellum. Three rings of the basal body are embedded in the cytoplasmic membrane, peptidoglycan layer and outer membrane. The outer filament is connected to the basal body. The cytoplasmic membrane ring of the basal body is associated with the Mot complex. This complex functions like a motor, rotating the flagellum thus rendering the cell motile. Energy for this rotation is provided from the proton (or sodium) motive force.

(b) The archaeal flagellum. The filament is composed of multiple proteins and the hook cannot be distinguished. ATP provides energy for flagellar rotation.

HAPI and HAP3, hook-associated proteins; HAP2, filament cap.

Springer, with permission.

The hook connects the central rod of the basal body to the filament and is composed of a single protein called the hook protein. The filament, with a diameter of 10–20 nm, can be dissolved at pH 3–4 with surfactants to a single protein solution of flagellin. The molecular weight of flagellin varies from 20 to 65 kD depending on the bacterial species. The hook and the filament are tube-shaped

and the flagellin moves through the tube to the growing tip of the filament. The tip of the filament is covered with filament cap protein. Flagellin can be exported to the medium in mutants defective in expression of this protein.

The number and location of flagella vary depending on the bacterial species. In some prokaryotes they are located at one or both poles, Cambridge University Press & Assessment 978-1-316-62291-9 — Prokaryotic Metabolism and Physiology Byung Hong Kim , Geoffrey Michael Gadd Excerpt <u>More Information</u>

10 | COMPOSITION AND STRUCTURE OF PROKARYOTIC CELLS

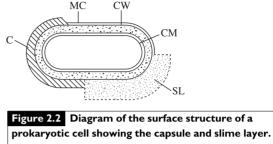
while the entire cell surface may be covered with flagella in others.

Although archaeal flagella resemble bacterial flagella in shape, they differ in composition and structure. The archaeal flagella are thinner than bacterial flagella, and composed of multiple proteins forming solid rods. The hook and filament cannot be distinguished, and the basal body is not in a ring form (Figure 2.1b). ATP is consumed to rotate archaeal flagella.

The fimbria, also known as the pilus (plural, pili), is observed in many Gram-negative bacteria but rarely in Gram-positive bacteria. Fimbriae have been proposed as the fibrils that mediate attachment to surfaces. For this reason, the term pilus should be used only to describe the F-pilus, the structure that mediates conjugation. Fimbriae are generally smaller in length $(0.2-20 \mu m)$ and width (3-14 nm) than flagella. Fimbriae help the organism to stick to surfaces of other bacteria, to host cells of animals and plants, and to solid surfaces. Different kinds of fimbriae are known which depend on the species as well as the growth conditions for a given organism. Fimbriae consist of a major protein with minor proteins called adhesins that facilitate bacterial attachment to surfaces by recognizing the appropriate receptor molecules. They are classified as type I through to type IV according to these receptor recognition properties. Adhesive properties are inhibited by sugars such as mannose, galactose and their oligomers, suggesting that the receptors are carbohydrate in nature.

A fibril bigger in size than fimbriae occurs in many Gram-negative bacteria that harbour the conjugative F-plasmid. This is called the F-pilus or sex pilus and mediates attachment between mating cells for the purpose of transmitting DNA from the donor cell by means of the F-pilus to a recipient cell. The F-pilus recognizes a receptor molecule on the surface of the recipient cell and after attachment, the F-pilus is depolymerized so that there is direct contact between the cells for DNA transmission.

Shewanella oneidensis and Geobacter sulfurreducens produce electrically conductive pilus-like appendages called bacterial nanowires, when



prokaryotic cell showing the capsule and slime layer. C, capsule; CM, cytoplasmic membrane; CW, cell wall; MC,

micro-capsule; SL, slime layer.

Fe(III) is used as the electron acceptor. These appendages are widespread in bacteria and are not exclusive to dissimilatory metal-reducing bacteria (Section 9.2.1). They may, in fact, reprecommon bacterial strategy sent а for efficient electron transfer and energy distribution. Similarly a micro-cable located in the periplasm of a bacterial strain belonging to the family Desulfobulbaceae transfers electrons through centimetre-long filaments spanning the aerobic surface to the anaerobic sulfide-rich zone of marine sediments (Section 10.3.1).

2.3.2 Capsules and slime layers

Many prokaryotic cells are covered with polysaccharides known as extracellular polymeric substances (EPS). In some cases the polymers are tightly integrated with the cell, while in others they are loosely associated. The former is called a capsule, and the latter a slime layer (Figure 2.2). Slime layer materials can diffuse into the medium, their structure and composition being dependent on growth conditions. An important role for these structures is adhesion to host cells for invasion or to a solid surface to initiate and stabilize biofilm formation. These structures are also responsible for resistance to phagocytosis, thereby increasing virulence. In some bacteria the capsule functions as a receptor for phage. Since the polysaccharides are hydrophilic, they can also protect cells from desiccation.

The term glycocalyx can be used to describe extracellular structures including the capsule and S-layer, the latter being described below.